

Neurogenesis in Human Hippocampus: Implications for Alzheimer Disease Pathogenesis

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Key Words

Alzheimer disease · Cell cycle · Hippocampal neurogenesis

Abstract

Hippocampal neurogenesis, the innate capacity of stem cells in the hippocampus to generate new neurons throughout life, is attracting interest in neurodegenerative disease research as an indicator of neuroplasticity and therefore treatment. Conditions that improve cognitive output and increase neurogenesis are associated with a decreased incidence of Alzheimer disease (AD), and conditions that lead to reduced cognition and neurogenesis are associated with increases in the incidence of AD. Therefore, hippocampal neurogenesis may be of particular relevance in the development of AD, and the modulation of this process can afford an important therapeutic avenue. Nevertheless, apparent contradictions across studies examining all factors of hippocampal neurogenesis in AD have led to a confusing state of affairs regarding the role of this process in the disease, as aberrant cell cycle activation has been advanced as an early pathogenic factor in the development of AD. The objective of this review is to critically examine these contradicting reports and provide additional and alternative insights into the function of hippocampal neurogenesis in AD.

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Alzheimer disease (AD) is the most common cause of dementia, and its prevalence is directly related to age [1]. Moreover, the increasing life expectancy, rising from 50 years at the beginning of the last century to approximately 80 years today, inevitably brings a higher incidence of age-related illnesses, including AD, resulting in an increase from 4 million individuals currently affected with AD in the United States to an estimated 14 million by 2050 [2]. In spite of these figures, the lack of progress toward effective therapy is striking.

The hippocampus is among the brain regions affected by AD pathology, particularly neurofibrillary pathology, early in disease [3, 4], a finding that is congruous with the early loss of spatial and episodic memory in AD [5]. Interestingly, the hippocampus is also one of few brain regions capable of neurogenesis in adulthood [6–8]. This property, in conjunction with the potential for exploiting regenerative capabilities of stem cells, offers some hope for a therapeutic intervention distinct from lesion-based therapies that have been disappointing to date.

Hippocampal Neurogenesis

Adult neurons are, in general, terminally differentiated and exist throughout the life of the organism. However, we now know that the dentate gyrus of the hippo-

campus [6–8], the olfactory bulb [8, 9] and certain areas of the cortex [10–14] retain the capacity to produce neurons during adulthood and that such neurons can integrate into the brain regions, organize, differentiate and become functional. In the adult dentate gyrus, cells divide continuously in the subgranular zone, located between the granular cell layer and the hilus of the hippocampus [15, 16]. Using the reagent bromodeoxyuridine (BrdU), which by incorporating into the DNA of dividing cells allows the identification of these cells in a time-dependent and location-specific manner, hippocampal cells have been shown to divide daily, and after a period of 4 weeks, 67% of the initially counted cells remain. In addition, of these remaining cells, about 50% continue to develop a neuronal phenotype and migrate to the granule cell layer, 15% differentiate into glial cells and the remaining 35% do not differentiate [17].

In adulthood, neurogenesis occurs in a variety of vertebrate species [18–21] including humans [22], and recent work suggests that newly generated granular neurons extend axons and make connections with the hippocampal CA3 region [23–25], receive synaptic input [23] and demonstrate functional properties indistinguishable from mature granule cells *in vivo* [26] and *in vitro* [25]. Therefore, neurons of the dentate gyrus of the hippocampus in adulthood can survive, differentiate into mature phenotypes and acquire functional properties indistinguishable from the mature cells of the region. As such, it is reasonable to assume that neurogenesis and the intrinsic capacity of the brain to generate new neurons may play a fundamental role in the various functional aspects of the hippocampus, including cognitive behavior.

Functional Significance

The mechanisms associated with hippocampal neurogenesis and its function and significance to learning and memory are presently being established. Thus far, it has been suggested that neurogenesis sustains the capacity of the dentate gyrus to maintain the continued modulation of cortical input supplied with novel complexities [27]. Interestingly, a substantial reduction in the number of newly generated neurons directly impairs hippocampal-dependent trace conditioning but had no effect on learning during hippocampal-independent tasks, suggesting a direct role for these cells in hippocampal-related memory [28].

Importantly, and as would be expected by the nature of hippocampal memory function, this process is also

highly susceptible to environmental/experience-dependent structural changes such that a number of factors can regulate the production and survival of hippocampal neurons developed during rodent adulthood. Factors such as estrogen [29–31], environmental complexity [18, 19, 32–36], exercise [37–39], N-methyl-D-aspartate-related excitatory input [40–42], transient global and focal ischemia [43–46] and epileptic seizures/kindling [47–53] positively regulate neurogenesis. Others such as depression [54–56], stress [20, 57–62], cholinergic denervation [63] and aging [15, 33, 36, 64] decrease the levels of neurogenesis.

Importantly, factors that affect neurogenesis correlate with changes in behavioral performance, further indicating a direct influence of neurogenesis on learning and memory [27, 28, 65–69]. For example, placing rodents in an enriched housing environment [18, 19, 32–36] or undergoing exercise [37–39] increases neurogenesis by increasing the survival of progenitor cells, leading to enhanced performance in the water maze task. Conversely, other factors such as stress, which is linked to a decrease in neurogenesis, impair cognitive performance [20, 57–62]. This involvement of hippocampal neurogenesis in the modulation of hippocampally related cognitive output suggests a potentially important role of this process in degenerative diseases which are associated with severe damage to the hippocampus, the most noteworthy being AD.

Neurogenesis in AD

The process of hippocampal neurogenesis as a potential therapeutic and also a pathogenic mechanism can be applied to the study of AD. In this regard, the positive modulators of neurogenesis and hippocampally controlled memory such as enriched environment [18, 19, 32–36], learning [27, 28, 65–69] and estrogen levels [29–31] have all been associated with a decreased incidence of AD [70–76]. Likewise, factors associated with decreased cognitive output and reduced neurogenesis, such as depression [54–56], stress [20, 57–62], cholinergic denervation [63, 77] and, most importantly, aging [15, 33, 36, 64], are all associated with an increased incidence of AD [1, 78–82]. These striking parallels suggest that AD may be associated with declines in hippocampal neurogenesis. As such, one obvious place to begin examining the possibility that hippocampal neurogenesis is directly associated with AD pathogenesis is by exploring the role of this process in the established animal models of this disease.

Neurogenesis and Animal Models of AD

In this regard, linkage studies show that mutations in at least 3 genes, the amyloid- β protein precursor (A β PP) gene located on chromosome 21 and the 2 homologous genes presenilin 1 and 2 (PS1 and PS2) located on chromosome 14 and 1, respectively, are associated with early-onset AD [83]. Based on these findings, several transgenic mouse models of AD have been generated to mimic the most extensively studied pathological hallmark of AD, namely amyloid- β plaques [84]. The creation of transgenic lines expressing mutated A β PP, a type 1 membrane protein with a large extracellular domain and a short cytoplasmic domain belonging to a larger A β PP family that also includes amyloid precursor-like proteins 1 and 2 [85], or mutations in the genes encoding PS1 and PS2, which are polytopic proteins with 6–8 transmembrane domains that play a central role in intramembrane proteolysis (δ -cleavage) of a number of cell surface proteins including A β PP and that give rise to amyloid- β peptide [86], have been used to establish these models as they share the common feature that they all alter γ -secretase cleavage of A β PP to increase the production of amyloid- β_{42} , the primary component of amyloid plaques. Likewise, the fact that the penetrance of the mutations is almost 100% suggests that they play critical roles in neurodegeneration and perhaps, given the parallels between modulators of neurogenesis and incidence factors in AD, these mutations/the overexpression of amyloid- β can also affect neurogenesis.

Two groups of investigators using transgenic animals exhibiting mutations in the A β PP have demonstrated declines in neurogenesis affecting both proliferation and survival of precursor cells [87–89]. Dong and colleagues [89] show declines in neurogenesis as early as 3 months, well before the deposition of amyloid- β plaques begins, suggesting that declines in neurogenesis may be independent of amyloid- β senile plaque formation. However, Haughey and colleagues [87] found that the declines in neurogenesis are restricted to the period when amyloid- β plaque deposition begins (12–14 months), and not in younger animals, suggesting an association with plaque deposition.

In addition to findings in A β PP transgenic animal models of AD, 3 studies in animals expressing the PS1 familial AD mutation also show impaired adult neurogenesis as measured by declines in precursor proliferation [90, 91] and survival [91], as well as various degrees of memory impairment [90, 91]. However, these differences are only visible when the animals are placed in an

enriched environment, an intervention that would maximize differences as it is a powerful positive modulator of neurogenesis. Even with the apparent perplexities across studies, research suggests that in AD, whether prior to amyloid- β deposition or during later stages of the disease when amyloid- β deposition is extensive, declines in hippocampal neurogenesis could be a factor related to hippocampal-associated learning and memory deficits readily observed in this disease.

Hippocampal Neurogenesis and Repair

Recent studies suggest a self-repair mechanism to be responsible for increases rather than decreases in hippocampal neurogenesis in AD. Several studies demonstrate that factors such as ischemia [43–46] or seizure-induced brain insults [47–53] lead to increased neurogenesis in the absence of improvements in cognition. Thus, one consideration is that neurogenesis may be activated as an endogenous repair mechanism utilized by the brain to diminish damage from trauma, blood-flow deprivation insults and/or overactivity [53]. This capacity of the brain to seemingly repair itself from these insults has been proposed as major grounds for the use of stem cells in AD [92–94]. To this end, the accumulation of pathological agents like amyloid- β or phosphorylated tau, which are in most cases described as toxic [95, 96], could potentially activate this self-repair mechanism. Even more significant is the fact that both brain trauma and ischemic episodes are associated with an increased incidence of AD. Therefore, the hypothesis that AD-related pathology, similar to ischemia or brain trauma, could drive the brain to attempt to self-regenerate by inducing neurogenesis is now gaining importance. However, based on this assumption, AD would not lead to reduced neurogenesis, as previously described, but to enhanced neurogenesis as suggested in the literature summarized below.

Modulation of Hippocampal Neurogenesis by AD Pathology

Invoking the brain repair theory has permitted investigators to explain the seemingly paradoxical findings concerning the presence of increased neurogenesis in AD [97], in animal models of the disease [98], and after *in vitro* treatment with amyloid- β [99]. In the latter, the administration of different concentrations of the amyloid- β peptide to neural stem cell cultures of hippocampi from

postnatal day 0 Bl6 mice leads to dose-dependent increases in the total number of cells [99]. Nevertheless, this report contradicts a previous study where treatment of neural stem cells with amyloid- β led to reduced neurogenesis and increased apoptosis of these cells [88]. Postnatal hippocampal cultures which include mature neurons in addition to glia, as those used in the Lopez-Toledano and Shelanski study [99], more closely resemble the hippocampal environment in vivo. Yet the use of amyloid- β_{42} , the peptide produced in the animal models of the disease and also used in the Lopez-Toledano and Shelanski study [99], did not mimic the findings observed in the animal models. Interestingly, in vitro studies have demonstrated that A β PP and its proteolytic fragments (i.e. amyloid- β peptide and secreted A β PP) have mitogenic properties [100–104]; hence, these results may merely reflect an in vitro specific event. Corroboration for the in vitro data is found in a recent study demonstrating that there is increased hippocampal neurogenesis in the brains of AD patients, as well as increased numbers of immature but not mature neuronal markers when compared with age-matched control brains [97]. Perplexingly, another study using animals carrying the Indiana and Swedish versions of the A β PP mutation has recently demonstrated that both young (3 months old) and older (12 months old) transgenic mice show enhanced BrdU labeling in the dentate gyrus [98], directly contradicting the previously mentioned reports [87–89]. These findings point to a possible repair-driven strategy of the AD brain to protect itself from pathology-associated damage. However, one possibility that remains unexplored, with regard to human studies, is that the brain may be attempting self-repair from the ischemic-like insults that occur late in the course of the disease [105], as mentioned before, a factor demonstrated to result in increased neurogenesis [43–46].

Oxidative Stress as a Modulator of Neurogenesis

While amyloid toxicity can be used to explain both increases and decreases in neurogenesis, it cannot simultaneously explain both, and hence, the confusion as to whether increases or decreases in neurogenesis exist in AD remains unresolved. Other processes are tightly associated with AD and can affect neurogenesis multi-dimensionally, and one such factor is oxidative stress. That hippocampal neurogenesis increases fail to yield any significant improvements in cognitive output points to the fact that mechanisms independent of amyloid deposition,

whether it is direct toxicity or toxicity-induced self-repair, may be at play. Evidence that the toxic nature of amyloid- β may not play a role in the modulation of neurogenesis stems from the fact that environmental manipulations that maximize hippocampal neurogenesis decrease amyloid- β load [106]. Since previous studies have reported that declines in neurogenesis were only evident in animals that were environmentally enriched [90, 91], when amyloid- β presumably decreases, amyloid- β toxicity does not seem to be the cause of neurogenesis declines.

One modulatory mechanism that can explain this confusing state of affairs, which is a key player in the pathogenesis of AD [107–112] and also a modulator of hippocampal neurogenesis [113–119], is oxidative stress.

In this regard, several reports demonstrate that antioxidant treatment [113–117] or antioxidant-related mechanisms such as caloric restriction [118] promote hippocampal neurogenesis, since oxidative stress precedes the deposition of amyloid- β in the AD brain [112], leading to the examination of an antioxidant role for amyloid- β [120–122]. It is the antioxidant role of amyloid- β that can indeed explain the perplexing results found in the AD neurogenesis literature.

In humans, the deposition of amyloid- β is associated with decreases in oxidative stress; therefore, it would not be the self-repair strategy induced by amyloid- β toxicity that leads to the promotion of neurogenesis in the AD brain but rather its antioxidant function. This could explain the increases in neurogenesis found in AD patients [97] and in cell models of the disease [99]. Thus, based on the above facts, this theory could not explain the declines in neurogenesis observed in animal models of the disease. However, when in excess or out of balance, amyloid- β , like all antioxidants, can act as a pro-oxidant, and hence, in the right circumstances, it can increase oxidative stress and lead to neurogenesis declines.

Finally, another possibility that should be considered is that a repair strategy is indeed implemented, not specifically by promoting hippocampal neurogenesis, but rather by using a parallel mechanism such as trying to reactivate the cell cycling machinery in vulnerable cell populations of the disease. It is well known that oxidative stress is a powerful modulator of the cell cycle and that oxidative stress conditions can lead to cell cycle aberration diseases such as cancer. This molecular event has now been suggested as a possible mechanism for AD pathogenesis and has been extensively reviewed [123–125]. In this regard, the absence of colabeling between BrdU and immature cell populations in the human AD

study [97] allows this theory to remain a possibility. Therefore, the multidimensional capacity of oxidative stress to modulate AD-related pathogenesis as well as cell cycle events establishes it as a viable model explaining AD-related increases and decreases in neurogenesis.

Contradictions

From the literature summarized above, the link between hippocampal neurogenesis and AD remains controversial. In addition to the contradictions in similar studies, many other inconsistencies further complicate this matter. For example, the fact that brain injury, which is associated with an increased incidence of AD, leads to increased hippocampal neurogenesis and that caloric restriction which increases neurogenesis [118], and is thought to be beneficial to normal subjects and to protect from many risk factors associated with AD (i.e. oxidative stress, inflammation), is detrimental or even lethal to the A β PP transgenic [126] contradicts this concept. More importantly, the fact that a recent report demonstrates that animal models of AD show decreased numbers of plaques under enriched environment [106], an intervention that increases neurogenesis and improves learning and memory [27, 28, 65–69], but yet other animal models of AD show that declines in neurogenesis are visible only under enriched environment conditions [90, 91], raises questions about the role of amyloid- β in neurogenesis. Moreover, one study demonstrating that aged animals with higher levels of new cells in the hippocampus show lower cognitive performance than those with fewer cells [127] only lends further confusion by suggesting that increases in neurogenesis, at times, may in fact represent a response to deleterious events and are not always associated with hippocampal ‘well-being.’

The findings presented in this review bring to light the inconsistencies within the field of hippocampal neurogenesis in general and in relation to AD and question the validity of the established animal models of this disease. In this regard, it is important to note that the genes associated with the mutations that lead to amyloid- β generation and that are used in these animal models also have powerful roles in neurodevelopment [128]. For example, mutant mice lacking all 3 A β PP genes display early lethality as well as a high incidence of cortical dysplasia, suggesting a critical role in neural development [129]. Moreover, PS1 is involved in γ -cleavage of the Notch receptor, which plays a critical role in determining cell fate during early development [130] and is also involved in

degradation [131, 132] resulting in a PS1-deficient mouse with profound developmental abnormalities [133, 134]. Transgenic mice that express constitutively active β -catenin in neuroepithelial precursor cells develop enlarged brains, because a greater proportion of neural precursors re-enter the cell cycle after mitosis [135]. Therefore, since both A β PP and PS1 seem to be tightly involved in cell cycle regulation during development, they are also likely to play a role in hippocampal neurogenesis in animal models of AD independently of any AD-related event, namely overexpression of amyloid- β .

In conclusion, the above stated conflicts suggest that rather than critically examining the true role of hippocampal neurogenesis in a disease such as AD, the field seems to use ‘increases’ or ‘decreases’ in neurogenesis as a superficial ‘utensil’ to fit the hypothesis that amyloid- β is detrimental in AD. If neurogenesis is increased, it is because it is associated with self-repair after amyloid- β toxicity [99], and if it is decreased, it is associated with declining cognition due to amyloid- β deposition [87, 88]. However, the fact that no correlations exist between amyloid- β and neurogenesis in any of these studies, in addition to the other mitogenic or antioxidant properties of this molecule proposed in this review, both of which are capable of altering neurogenesis, makes such assumptions premature. Studies of neurogenesis in models other than APP mice, such as the NGF knockout or SAMP8 mouse, are warranted to further dissect the relationship between adult neurogenesis and AD. Likewise, studies directly targeting hippocampal neurogenesis in normal mice, using AD-associated markers such as amyloid- β infusion, would further help elucidating a direct role between AD, amyloid- β and neurogenesis. Therefore, to date, no compensation or crisis, but rather a chaotic state of affairs seems to be present in the literature regarding the role of neurogenesis in AD, and, judging from the disparity in results across the neurogenesis-Alzheimer studies, the jury is still out on the exact function of neurogenesis.

References

- 1 Katzman R: Alzheimer's disease as an age-dependent disorder. *Ciba Found Symp* 1988; 134:69–85.
- 2 Larson EB, Kukull WA, Katzman RL: Cognitive impairment: dementia and Alzheimer's disease. *Annu Rev Public Health* 1992;13: 431–449.
- 3 Avila J, Lim F, Moreno F, Belmonte C, Cuello AC: Tau function and dysfunction in neurons: its role in neurodegenerative disorders. *Mol Neurobiol* 2002;25:213–231.

- 4 Selkoe DJ: Images in neuroscience. Alzheimer's disease: from genes to pathogenesis. *Am J Psychiatry* 1997;154:1198.
- 5 de Toledo-Morrell L, Dickerson B, Sullivan MP, Spanovic C, Wilson R, Bennett DA: Hemispheric differences in hippocampal volume predict verbal and spatial memory performance in patients with Alzheimer's disease. *Hippocampus* 2000;10:136-142.
- 6 Altman J, Das GD: Post-natal origin of microneurons in the rat brain. *Nature* 1965;207:953-956.
- 7 Rakic P: DNA synthesis and cell division in the adult primate brain. *Ann N Y Acad Sci* 1985;457:193-211.
- 8 Suhonen JO, Peterson DA, Ray J, Gage FH: Differentiation of adult hippocampus-derived progenitors into olfactory neurons in vivo. *Nature* 1996;383:624-627.
- 9 Luskin MB: Restricted proliferation and migration of postnatally generated neurons derived from the forebrain subventricular zone. *Neuron* 1993;11:173-189.
- 10 Kriegstein AR: Cortical neurogenesis and its disorders. *Curr Opin Neurol* 1996;9:113-117.
- 11 Perez-Canellas MM, Garcia-Verdugo JM: Adult neurogenesis in the telencephalon of a lizard: a [3H]thymidine autoradiographic and bromodeoxyuridine immunocytochemical study. *Brain Res Dev Brain Res* 1996;93:49-61.
- 12 Gould E, Reeves AJ, Graziano MS, Gross CG: Neurogenesis in the neocortex of adult primates. *Science* 1999;286:548-552.
- 13 Magavi SS, Macklis JD: Induction of neuronal type-specific neurogenesis in the cerebral cortex of adult mice: manipulation of neural precursors in situ. *Brain Res Dev Brain Res* 2002;134:57-76.
- 14 Bayer SA, Altman J: Hippocampal development in the rat: cytogenesis and morphogenesis examined with autoradiography and low-level X-irradiation. *J Comp Neurol* 1974;158:55-79.
- 15 Kuhn HG, Dickinson-Anson H, Gage FH: Neurogenesis in the dentate gyrus of the adult rat: age-related decrease of neuronal progenitor proliferation. *J Neurosci* 1996;16:2027-2033.
- 16 Gage FH, Kempermann G, Palmer TD, Peterson DA, Ray J: Multipotent progenitor cells in the adult dentate gyrus. *J Neurobiol* 1998;36:249-266.
- 17 Kempermann G, Kuhn HG, Gage FH: Genetic influence on neurogenesis in the dentate gyrus of adult mice. *Proc Natl Acad Sci USA* 1997;94:10409-10414.
- 18 Kempermann G, Brandon EP, Gage FH: Environmental stimulation of 129/SvJ mice causes increased cell proliferation and neurogenesis in the adult dentate gyrus. *Curr Biol* 1998;8:939-942.
- 19 Nilsson M, Perfilieva E, Johansson U, Orwar O, Eriksson PS: Enriched environment increases neurogenesis in the adult rat dentate gyrus and improves spatial memory. *J Neurobiol* 1999;39:569-578.
- 20 Gould E, McEwen BS, Tanapat P, Galea LA, Fuchs E: Neurogenesis in the dentate gyrus of the adult tree shrew is regulated by psychosocial stress and NMDA receptor activation. *J Neurosci* 1997;17:2492-2498.
- 21 Gould E, Reeves AJ, Fallah M, Tanapat P, Gross CG, Fuchs E: Hippocampal neurogenesis in adult Old World primates. *Proc Natl Acad Sci USA* 1999;96:5263-5267.
- 22 Eriksson PS, Perfilieva E, Bjork-Eriksson T, Alborn AM, Nordborg C, Peterson DA, Gage FH: Neurogenesis in the adult human hippocampus. *Nat Med* 1998;4:1313-1317.
- 23 Markakis EA, Gage FH: Adult-generated neurons in the dentate gyrus send axonal projections to field CA3 and are surrounded by synaptic vesicles. *J Comp Neurol* 1999;406:449-460.
- 24 Hastings NB, Gould E: Rapid extension of axons into the CA3 region by adult-generated granule cells. *J Comp Neurol* 1999;413:146-154.
- 25 Song HJ, Stevens CF, Gage FH: Neural stem cells from adult hippocampus develop essential properties of functional CNS neurons. *Nat Neurosci* 2002;5:438-445.
- 26 van Praag H, Schinder AF, Christie BR, Toni N, Palmer TD, Gage FH: Functional neurogenesis in the adult hippocampus. *Nature* 2002;415:1030-1034.
- 27 Kempermann G: Why new neurons? Possible functions for adult hippocampal neurogenesis. *J Neurosci* 2002;22:635-638.
- 28 Shors TJ, Miesegaes G, Beylin A, Zhao M, Rydel T, Gould E: Neurogenesis in the adult is involved in the formation of trace memories. *Nature* 2001;410:372-376.
- 29 Tanapat P, Hastings NB, Reeves AJ, Gould E: Estrogen stimulates a transient increase in the number of new neurons in the dentate gyrus of the adult female rat. *J Neurosci* 1999;19:5792-5801.
- 30 Saravia F, Reysin Y, Lux-Lantos V, Beauquis J, Homo-Delarche F, De Nicola AF: Oestradiol restores cell proliferation in dentate gyrus and subventricular zone of streptozotocin-diabetic mice. *J Neuroendocrinol* 2004;16:704-710.
- 31 Perez-Martin M, Azcoitia I, Trejo JL, Sierra A, Garcia-Segura LM: An antagonist of estrogen receptors blocks the induction of adult neurogenesis by insulin-like growth factor-I in the dentate gyrus of adult female rat. *Eur J Neurosci* 2003;18:923-930.
- 32 Brown J, Cooper-Kuhn CM, Kempermann G, Van Praag H, Winkler J, Gage FH, Kuhn HG: Enriched environment and physical activity stimulate hippocampal but not olfactory bulb neurogenesis. *Eur J Neurosci* 2003;17:2042-2046.
- 33 Kempermann G, Gast D, Gage FH: Neuroplasticity in old age: sustained fivefold induction of hippocampal neurogenesis by long-term environmental enrichment. *Ann Neurol* 2002;52:135-143.
- 34 Kempermann G, Gage FH: Experience-dependent regulation of adult hippocampal neurogenesis: effects of long-term stimulation and stimulus withdrawal. *Hippocampus* 1999;9:321-332.
- 35 Kempermann G, Kuhn HG, Gage FH: More hippocampal neurons in adult mice living in an enriched environment. *Nature* 1997;386:493-495.
- 36 Kempermann G, Kuhn HG, Gage FH: Experience-induced neurogenesis in the senescent dentate gyrus. *J Neurosci* 1998;18:3206-3212.
- 37 Kempermann G, van Praag H, Gage FH: Activity-dependent regulation of neuronal plasticity and self repair. *Prog Brain Res* 2000;127:35-48.
- 38 van Praag H, Kempermann G, Gage FH: Running increases cell proliferation and neurogenesis in the adult mouse dentate gyrus. *Nat Neurosci* 1999;2:266-270.
- 39 Kitamura T, Mishina M, Sugiyama H: Enhancement of neurogenesis by running wheel exercises is suppressed in mice lacking NMDA receptor epsilon1 subunit. *Neurosci Res* 2003;47:55-63.
- 40 Arvidsson A, Kokaia Z, Lindvall O: N-methyl-D-aspartate receptor-mediated increase of neurogenesis in adult rat dentate gyrus following stroke. *Eur J Neurosci* 2001;14:10-18.
- 41 Cameron HA, McEwen BS, Gould E: Regulation of adult neurogenesis by excitatory input and NMDA receptor activation in the dentate gyrus. *J Neurosci* 1995;15:4687-4692.
- 42 Nacher J, Rosell DR, Alonso-Llosa G, McEwen BS: NMDA receptor antagonist treatment induces a long-lasting increase in the number of proliferating cells, PSA-NCAM-immunoreactive granule neurons and radial glia in the adult rat dentate gyrus. *Eur J Neurosci* 2001;13:512-520.
- 43 Yagita Y, Kitagawa K, Ohtsuki T, Takasawa K, Miyata T, Okano H, Hori M, Matsumoto M: Neurogenesis by progenitor cells in the ischemic adult rat hippocampus. *Stroke* 2001;32:1890-1896.
- 44 Liu J, Solway K, Messing RO, Sharp FR: Increased neurogenesis in the dentate gyrus after transient global ischemia in gerbils. *J Neurosci* 1998;18:7768-7778.
- 45 Kee NJ, Preston E, Wojtowicz JM: Enhanced neurogenesis after transient global ischemia in the dentate gyrus of the rat. *Exp Brain Res* 2001;136:313-320.
- 46 Jin K, Minami M, Lan JQ, Mao XO, Bateur S, Simon RP, Greenberg DA: Neurogenesis in dentate subgranular zone and rostral subventricular zone after focal cerebral ischemia in the rat. *Proc Natl Acad Sci USA* 2001;98:4710-4715.
- 47 Parent JM, Yu TW, Leibowitz RT, Geschwind DH, Sloviter RS, Lowenstein DH: Dentate granule cell neurogenesis is increased by seizures and contributes to aberrant network reorganization in the adult rat hippocampus. *J Neurosci* 1997;17:3727-3738.
- 48 Scott BW, Wojtowicz JM, Burnham WM: Neurogenesis in the dentate gyrus of the rat following electroconvulsive shock seizures. *Exp Neurol* 2000;165:231-236.

- 49 Wang YL, Sun RP, Lei GF, Wang JW, Guo SH: Neurogenesis of dentate granule cells following kainic acid induced seizures in immature rats. *Zhonghua Er Ke Za Zhi* 2004; 42:621–624.
- 50 Cha BH, Akman C, Silveira DC, Liu X, Holmes GL: Spontaneous recurrent seizure following status epilepticus enhances dentate gyrus neurogenesis. *Brain Dev* 2004;26:394–397.
- 51 Mohapel P, Ekdahl CT, Lindvall O: Status epilepticus severity influences the long-term outcome of neurogenesis in the adult dentate gyrus. *Neurobiol Dis* 2004;15:196–205.
- 52 Hellsten J, Wennstrom M, Bengzon J, Mohapel P, Tingstrom A: Electroconvulsive seizures induce endothelial cell proliferation in adult rat hippocampus. *Biol Psychiatry* 2004; 55:420–427.
- 53 Parent JM: The role of seizure-induced neurogenesis in epileptogenesis and brain repair. *Epilepsy Res* 2002;50:179–189.
- 54 Malberg JE, Eisch AJ, Nestler EJ, Duman RS: Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. *J Neurosci* 2000;20:9104–9110.
- 55 Malberg JE, Duman RS: Cell proliferation in adult hippocampus is decreased by inescapable stress: reversal by fluoxetine treatment. *Neuropsychopharmacology* 2003;28:1562–1571.
- 56 Jacobs BL: Adult brain neurogenesis and depression. *Brain Behav Immun* 2002;16:602–609.
- 57 Tanapat P, Galea LA, Gould E: Stress inhibits the proliferation of granule cell precursors in the developing dentate gyrus. *Int J Dev Neurosci* 1998;16:235–239.
- 58 McEwen BS: Stress and hippocampal plasticity. *Annu Rev Neurosci* 1999;22:105–122.
- 59 Czeh B, Welt T, Fischer AK, Erhardt A, Schmitt W, Muller MB, Toschi N, Fuchs E, Keck ME: Chronic psychosocial stress and concomitant repetitive transcranial magnetic stimulation: effects on stress hormone levels and adult hippocampal neurogenesis. *Biol Psychiatry* 2002;52:1057–1065.
- 60 Pham K, Nacher J, Hof PR, McEwen BS: Repeated restraint stress suppresses neurogenesis and induces biphasic PSA-NCAM expression in the adult rat dentate gyrus. *Eur J Neurosci* 2003;17:879–886.
- 61 Lemaire V, Koehl M, Le Moal M, Abrous DN: Prenatal stress produces learning deficits associated with an inhibition of neurogenesis in the hippocampus. *Proc Natl Acad Sci USA* 2000;97:11032–11037.
- 62 Lemaire V, Aurousseau C, Le Moal M, Abrous DN: Behavioural trait of reactivity to novelty is related to hippocampal neurogenesis. *Eur J Neurosci* 1999;11:4006–4014.
- 63 Cooper-Kuhn CM, Winkler J, Kuhn HG: Decreased neurogenesis after cholinergic forebrain lesion in the adult rat. *J Neurosci Res* 2004;77:155–165.
- 64 Bizon JL, Gallagher M: Production of new cells in the rat dentate gyrus over the lifespan: relation to cognitive decline. *Eur J Neurosci* 2003;18:215–219.
- 65 Gould E, Tanapat P, Hastings NB, Shors TJ: Neurogenesis in adulthood: a possible role in learning. *Trends Cogn Sci* 1999;3:186–192.
- 66 Gould E, Beylin A, Tanapat P, Reeves A, Shors TJ: Learning enhances adult neurogenesis in the hippocampal formation. *Nat Neurosci* 1999;2:260–265.
- 67 Drapeau E, Mayo W, Aurousseau C, Le Moal M, Piazza PV, Abrous DN: Spatial memory performances of aged rats in the water maze predict levels of hippocampal neurogenesis. *Proc Natl Acad Sci USA* 2003;100:14385–14390.
- 68 Prickaerts J, Koopmans G, Blokland A, Scheepens A: Learning and adult neurogenesis: survival with or without proliferation? *Neurobiol Learn Mem* 2004;81:1–11.
- 69 Schmidt-Hieber C, Jonas P, Bischofberger J: Enhanced synaptic plasticity in newly generated granule cells of the adult hippocampus. *Nature* 2004;429:184–187.
- 70 Pope SK, Shue VM, Beck C: Will a healthy lifestyle help prevent Alzheimer's disease? *Annu Rev Public Health* 2003;24:111–132.
- 71 Callahan CM, Hall KS, Hui SL, Musick BS, Unverzagt FW, Hendrie HC: Relationship of age, education, and occupation with dementia among a community-based sample of African Americans. *Arch Neurol* 1996;53:134–140.
- 72 Glatt SL, Hubble JP, Lyons K, Paolo A, Troster AI, Hassanein RE, Koller WC: Risk factors for dementia in Parkinson's disease: effect of education. *Neuroepidemiology* 1996;15:20–25.
- 73 Fritsch T, McClendon MJ, Smyth KA, Ogrocki PK: Effects of educational attainment and occupational status on cognitive and functional decline in persons with Alzheimer-type dementia. *Int Psychogeriatr* 2002;14:347–363.
- 74 Tang MX, Jacobs D, Stern Y, Marder K, Schofield P, Gurland B, Andrews H, Mayeux R: Effect of oestrogen during menopause on risk and age at onset of Alzheimer's disease. *Lancet* 1996;348:429–432.
- 75 Stoppe G: Dementia: risk and protective factors with special consideration of gender and hormone replacement therapy. *Z Arztl Fortbild Qualitatssich* 2000;94:217–222.
- 76 Geerlings MI, Ruitenberg A, Witteman JC, van Swieten JC, Hofman A, van Duijn CM, Breteler MM, Launer LJ: Reproductive period and risk of dementia in postmenopausal women. *JAMA* 2001;285:1475–1481.
- 77 Mohapel P, Leanza G, Kokaia M, Lindvall O: Forebrain acetylcholine regulates adult hippocampal neurogenesis and learning. *Neurobiol Aging* 2005;26:939–946.
- 78 Green RC, Cupples LA, Kurz A, Auerbach S, Go R, Sadovnick D, Duara R, Kukull WA, Chui H, Edeki T, Griffith PA, Friedland RP, Bachman D, Farrer L: Depression as a risk factor for Alzheimer disease: the MIRAGE Study. *Arch Neurol* 2003;60:753–759.
- 79 Speck CE, Kukull WA, Brenner DE, Bowen JD, McCormick WC, Teri L, Pfanschmidt ML, Thompson JD, Larson EB: History of depression as a risk factor for Alzheimer's disease. *Epidemiology* 1995;6:366–369.
- 80 Wilson RS, Evans DA, Bienias JL, Mendes de Leon CF, Schneider JA, Bennett DA: Prone to psychological distress is associated with risk of Alzheimer's disease. *Neurology* 2003;61:1479–1485.
- 81 Aronson MK, Ooi WL, Geva DL, Masur D, Blau A, Frishman W: Dementia. Age-dependent incidence, prevalence, and mortality in the old. *Arch Intern Med* 1991;151:989–992.
- 82 Gavrilova SI, Bratsun AL: Epidemiology and risk factors of Alzheimer's disease. *Vestn Ross Akad Med Nauk* 1999;39–46.
- 83 Hardy J: Amyloid, the presenilins and Alzheimer's disease. *Trends Neurosci* 1997;20: 154–159.
- 84 Selkoe DJ: Alzheimer's disease: genes, proteins, and therapy. *Physiol Rev* 2001;81:741–766.
- 85 Coulson EJ, Paliga K, Beyreuther K, Masters CL: What the evolution of the amyloid protein precursor supergene family tells us about its function. *Neurochem Int* 2000;36: 175–184.
- 86 Selkoe DJ, Schenk D: Alzheimer's disease: molecular understanding predicts amyloid-based therapeutics. *Annu Rev Pharmacol Toxicol* 2003;43:545–584.
- 87 Haughey NJ, Liu D, Nath A, Borchard AC, Mattson MP: Disruption of neurogenesis in the subventricular zone of adult mice, and in human cortical neuronal precursor cells in culture, by amyloid beta-peptide: implications for the pathogenesis of Alzheimer's disease. *Neuromolecular Med* 2002;1:125–135.
- 88 Haughey NJ, Nath A, Chan SL, Borchard AC, Rao MS, Mattson MP: Disruption of neurogenesis by amyloid beta-peptide, and perturbed neural progenitor cell homeostasis, in models of Alzheimer's disease. *J Neurochem* 2002;83:1509–1524.
- 89 Dong H, Goico B, Martin M, Csernansky CA, Bertchume A, Csernansky JG: Modulation of hippocampal cell proliferation, memory, and amyloid plaque deposition in APPsw (Tg2576) mutant mice by isolation stress. *Neuroscience* 2004;127:601–609.
- 90 Feng R, Rampon C, Tang YP, Shrom D, Jin J, Kyin M, Sopher B, Miller MW, Ware CB, Martin GM, Kim SH, Langdon RB, Sisodia SS, Tsien JZ: Deficient neurogenesis in forebrain-specific presenilin-1 knockout mice is associated with reduced clearance of hippocampal memory traces. *Neuron* 2001;32: 911–926.
- 91 Wang R, Dineley KT, Sweatt JD, Zheng H: Presenilin 1 familial Alzheimer's disease mutation leads to defective associative learning and impaired adult neurogenesis. *Neuroscience* 2004;126:305–312.
- 92 Gage FH: Brain, repair yourself. *Sci Am* 2003;289:46–53.
- 93 Reinoso Suarez F: Adult neurogenesis and stem cells. Functional capacity. *An R Acad Nac Med (Madr)* 2002;119:507–521; discussion 521–528.
- 94 Domanska-Janik K: Stem cells – potential therapeutic use in neurological diseases. *Neurol Neurochir Pol* 2002;36(suppl 1):107–117.

- 95 Bateman DA, Chakrabartty A: Interactions of Alzheimer amyloid peptides with cultured cells and brain tissue, and their biological consequences. *Biopolymers* 2004; 76:4–14.
- 96 Walsh DM, Klyubin I, Fadeeva JV, Rowan MJ, Selkoe DJ: Amyloid-beta oligomers: their production, toxicity and therapeutic inhibition. *Biochem Soc Trans* 2002;30: 552–557.
- 97 Jin K, Peel AL, Mao XO, Xie L, Cottrell BA, Henshall DC, Greenberg DA: Increased hippocampal neurogenesis in Alzheimer's disease. *Proc Natl Acad Sci USA* 2004;101: 343–347.
- 98 Jin K, Galvan V, Xie L, Mao XO, Gorostiza OF, Bredesen DE, Greenberg DA: Enhanced neurogenesis in Alzheimer's disease transgenic (PDGF-APP^{Sw,Ind}) mice. *Proc Natl Acad Sci USA* 2004;101:13363–13367.
- 99 Lopez-Toledano MA, Shelanski ML: Neurogenic effect of beta-amyloid peptide in the development of neural stem cells. *J Neurosci* 2004;24:5439–5444.
- 100 Milward EA, Papadopoulos R, Fuller SJ, Moir RD, Small D, Beyreuther K, Masters CL: The amyloid protein precursor of Alzheimer's disease is a mediator of the effects of nerve growth factor on neurite outgrowth. *Neuron* 1992;9:129–137.
- 101 Schubert D, Cole G, Saitoh T, Oltersdorf T: Amyloid beta protein precursor is a mitogen. *Biochem Biophys Res Commun* 1989; 162:83–88.
- 102 Copani A, Condorelli F, Canonico PL, Nicoletti F, Sortino MA: Cell cycle progression towards Alzheimer's disease. *Funct Neurol* 2001;16:11–15.
- 103 Hoffmann J, Twisselmann C, Kummer MP, Romagnoli P, Herzog V: A possible role for the Alzheimer amyloid precursor protein in the regulation of epidermal basal cell proliferation. *Eur J Cell Biol* 2000;79:905–914.
- 104 Schmitz A, Tikkanen R, Kirfel G, Herzog V: The biological role of the Alzheimer amyloid precursor protein in epithelial cells. *Histochem Cell Biol* 2002;117:171–180.
- 105 Jellinger KA, Mitter-Ferstl E: The impact of cerebrovascular lesions in Alzheimer disease – a comparative autopsy study. *J Neurol* 2003;250:1050–1055.
- 106 Lazarov O, Robinson J, Tang YP, Hairston IS, Korade-Mirnic Z, Lee VM, Hersh LB, Sapolsky RM, Mirnic K, Sisodia SS: Environmental enrichment reduces Aβ levels and amyloid deposition in transgenic mice. *Cell* 2005;120:701–713.
- 107 Smith MA, Sayre LM, Monnier VM, Perry G: Radical AGEing in Alzheimer's disease. *Trends Neurosci* 1995;18:172–176.
- 108 Smith MA, Perry G, Richey PL, Sayre LM, Anderson VE, Beal MF, Kowall N: Oxidative damage in Alzheimer's. *Nature* 1996; 382:120–121.
- 109 Perry G, Castellani RJ, Hirai K, Smith MA: Reactive oxygen species mediate cellular damage in Alzheimer disease. *J Alzheimers Dis* 1998;1:45–55.
- 110 Perry G, Smith MA: Is oxidative damage central to the pathogenesis of Alzheimer disease? *Acta Neurol Belg* 1998;98:175–179.
- 111 Smith MA, Rottkamp CA, Nunomura A, Raina AK, Perry G: Oxidative stress in Alzheimer's disease. *Biochim Biophys Acta* 2000;1502:139–144.
- 112 Nunomura A, Perry G, Aliev G, Hirai K, Takeda A, Balraj EK, Jones PK, Ghanbari H, Wataya T, Shimohama S, Chiba S, Atwood CS, Petersen RB, Smith MA: Oxidative damage is the earliest event in Alzheimer disease. *J Neuropathol Exp Neurol* 2001;60: 759–767.
- 113 Herrera DG, Yague AG, Johnsen-Soriano S, Bosch-Morell F, Collado-Morente L, Muriach M, Romero FJ, Garcia-Verdugo JM: Selective impairment of hippocampal neurogenesis by chronic alcoholism: protective effects of an antioxidant. *Proc Natl Acad Sci USA* 2003;100:7919–7924.
- 114 Cecchini T, Ciaroni S, Ferri P, Ambrogini P, Cuppini R, Santi S, Del Grande P: Alpha-tocopherol, an exogenous factor of adult hippocampal neurogenesis regulation. *J Neurosci Res* 2003;73:447–455.
- 115 Cuppini R, Ciaroni S, Cecchini T, Ambrogini P, Ferri P, Cuppini C, Ninfali P, Del Grande P: Tocopherols enhance neurogenesis in dentate gyrus of adult rats. *Int J Vitam Nutr Res* 2002;72:170–176.
- 116 Shen LH, Zhang JT: Ginsenoside Rg1 promotes proliferation of hippocampal progenitor cells. *Neurol Res* 2004;26:422–428.
- 117 Casadesus G, Shukitt-Hale B, Stellwagen HM, Zhu X, Lee HG, Smith MA, Joseph JA: Modulation of hippocampal plasticity and cognitive behavior by short-term blueberry supplementation in aged rats. *Nutr Neurosci* 2004;7:309–316.
- 118 Lee J, Duan W, Long JM, Ingram DK, Mattson MP: Dietary restriction increases the number of newly generated neural cells, and induces BDNF expression, in the dentate gyrus of rats. *J Mol Neurosci* 2000;15: 99–108.
- 119 Casadesus G, Shukitt-Hale B, Stellwagen HM, Smith MA, Rabin BM, Joseph JA: Hippocampal neurogenesis and PSA-NCAM expression following exposure to (56)Fe particles mimics that seen during aging in rats. *Exp Gerontol* 2005;40:249–254.
- 120 Joseph J, Shukitt-Hale B, Denisova NA, Martin A, Perry G, Smith MA: Copernicus revisited: amyloid beta in Alzheimer's disease. *Neurobiol Aging* 2001;22:131–146.
- 121 Rottkamp CA, Atwood CS, Joseph JA, Nunomura A, Perry G, Smith MA: The state versus amyloid-beta: the trial of the most wanted criminal in Alzheimer disease. *Peptides* 2002;23:1333–1341.
- 122 Smith MA, Atwood CS, Joseph JA, Perry G: Ill-fated amyloid-beta vaccine. *J Neurosci Res* 2002;69:285.
- 123 Raina AK, Zhu X, Rottkamp CA, Monteiro M, Takeda A, Smith MA: Cyclin' toward dementia: cell cycle abnormalities and abortive oncogenesis in Alzheimer disease. *J Neurosci Res* 2000;61:128–133.
- 124 Raina AK, Zhu X, Smith MA: Alzheimer's disease and the cell cycle. *Acta Neurobiol Exp (Wars)* 2004;64:107–112.
- 125 Zhu X, Raina AK, Perry G, Smith MA: Alzheimer's disease: the two-hit hypothesis. *Lancet Neurol* 2004;3:219–226.
- 126 Mattson MP: Neuroprotective signaling and the aging brain: take away my food and let me run. *Brain Res* 2000;886:47–53.
- 127 Bizon JL, Lee HJ, Gallagher M: Neurogenesis in a rat model of age-related cognitive decline. *Aging Cell* 2004;3:227–234.
- 128 Bothwell M, Giniger E: Alzheimer's disease: neurodevelopment converges with neurodegeneration. *Cell* 2000;102:271–273.
- 129 Koo EH: The beta-amyloid precursor protein (APP) and Alzheimer's disease: does the tail wag the dog? *Traffic* 2002;3:763–770.
- 130 Selkoe D, Kopan R: Notch and presenilin: regulated intramembrane proteolysis links development and degeneration. *Annu Rev Neurosci* 2003;26:565–597.
- 131 Kang DE, Soriano S, Xia X, Eberhart CG, De Strooper B, Zheng H, Koo EH: Presenilin couples the paired phosphorylation of beta-catenin independent of axin: implications for beta-catenin activation in tumorigenesis. *Cell* 2002;110:751–762.
- 132 Willert K, Nusse R: Beta-catenin: a key mediator of Wnt signaling. *Curr Opin Genet Dev* 1998;8:95–102.
- 133 Shen J, Bronson RT, Chen DF, Xia W, Selkoe DJ, Tonegawa S: Skeletal and CNS defects in presenilin-1-deficient mice. *Cell* 1997;89: 629–639.
- 134 Wong PC, Zheng H, Chen H, Becher MW, Sirinathsinghji DJ, Trumbauer ME, Chen HY, Price DL, Van der Ploeg LH, Sisodia SS: Presenilin 1 is required for Notch1 and Dll1 expression in the paraxial mesoderm. *Nature* 1997;387:288–292.
- 135 Chenn A, Walsh CA: Regulation of cerebral cortical size by control of cell cycle exit in neural precursors. *Science* 2002;297:365–369.